



Remediation Potential of Select Wetland Plants

Aparna Gunjal¹, Sudarshan Bhat² and T.V. Ramachandra^{2,*}

¹Department of Microbiology, Dr. D.Y. Patil Arts, Commerce and Science College, Pimpri, Pune, Maharashtra, India.

²Energy and Wetlands Research Group, CES T15, Indian Institute of Science, Bengaluru 560012, India.

* Email: tvr@iisc.ac.in

ABSTRACT

Phytoremediation refers to the ability of certain plants to bioaccumulate, degrade, or render harmless the contaminants in soil, water, etc. Wetlands function as the kidneys of a landscape as plants in wetlands act as sinks by uptake of nutrients and heavy metals. The current study focuses on estimating the phytoremediation potential of some selected wetland plants. Plant samples were collected from inlets and outlets of Jakkur wetlands, Bengaluru, India on a weekly basis. The plants were identified using standard keys based on taxonomic literature. Plants identified were *Typha sp.*, *Cyperus sp.*, *Ludwigia sp.*, *Pistia stratiotes*, *Polygonum sp.*, *Alternanthera philoxeroides* and *Spirodela sp.*, which are perennial. Among these, *Typha sp.*, *Cyperus sp.*, *Pistia stratiotes* and *Spirodela sp.* are monocots, and *Ludwigia sp.*, *Polygonum sp.* and *Alternanthera philoxeroides* are dicots. These plants showed the uptake of the heavy metals viz., cadmium, zinc, nickel, copper, chromium and lead. During the first week, uptake of Cu and Zn was more by young *Typha sp.* at inlet and Cr was more at outlet of sampling. The uptake of Cu, Pb, Zn and Ni was 1.4, 9.0, 9.4 and 10.0 mg/kg respectively during the second week of sampling in young *Typha sp.* shoot. Also, the Pb and Zn uptake was 13.6 and 10.4 mg/kg respectively in the mature *Typha sp.* during the third week at outlet of sampling. Ni uptake was 4.0 mg/kg during the third week of sampling by mature *Typha sp.* at inlet. The uptake of Pb, Zn and Cd was 12.2, 23.0 and 1.4 mg/kg respectively during the fourth week of sampling in the mature *Typha sp.* at outlet. The uptake of Pb, Ni, Cr and Cd was 5.2, 2.4, 12 and 0.8 mg/kg respectively during the first week of sampling in the medium *Polygonum sp.* The uptake of Zn was 10.2 mg/kg during the second week in the mature *Alternanthera philoxeroides* at outlet of sampling. The uptake of Cr was 19.6 mg/kg during the third week was found to be more by mature *Alternanthera philoxeroides* at outlet of sampling. The uptake of Cu, Pb, Zn and Cr was 2.2, 9.8, 8.6 and 10.2 mg/kg respectively during the fourth week in young *Alternanthera philoxeroides* at inlet. Similarly, the uptake of Cr was 13.6 mg/kg at outlet during the second week in *Spirodela sp.* The uptake of Cu was 2.4 mg/kg during the third week in medium *Ludwigia sp.* at outlet.

Keywords: Phytoremediation, Macrophytes, Physiology, Wetland, Nutrients, Perennial

Introduction

Metallic elements that have a high atomic weight, relatively high density and are toxic at low concentrations such as mercury (Hg), cadmium (Cd), arsenic (As), chromium (Cr), thallium (Tl), lead (Pb), etc. are referred

to as heavy metals. Applications of metals include domestic, agricultural, medical and technological with wider distribution in the environment. The potential effects on human health and the environment have been a concern, which necessitates suitable methods which can help in the remediation to minimise harmful effects. Soil and



water are increasingly getting contaminated with heavy metals¹ due to sustained flow of untreated effluents which is damaging the environment. Sources of heavy metals in the environment are geogenic, industrial, agricultural, pharmaceutical, domestic effluent, etc. Escalated health concerns associated with environmental contamination due to heavy metals have necessitated remediation measures that are economical and technically feasible. Remediation is to avoid all such environmental damage. Remediation can be achieved with the help of various physico-chemical processes viz., ion-exchange, precipitation, reverse osmosis, ultrafiltration, flocculation, electro dialysis, etc. But these methods are costly, time-consuming, and are also not very effective².

Bioremediation is a viable and cost effective approach using microorganisms (bacteria, actinobacteria, fungi, algae and yeasts), plant residues and plants to remove heavy metals from waste water, industrial effluents, etc.^{3,4}. Bioremediation is emerging as a green approach for the removal of heavy metals and other pollutants from contaminated land, water, etc.

Phytoremediation is one of the most eco-friendly, economical, simple and effective bioremediation options to remove heavy metals from waste water, industrial effluents, etc. In phytoremediation, the contaminated soils and water are treated *in-situ* with the help of plants⁵. Aquatic plants are of importance in this regard due to uptake mechanisms and rates of uptake of metals of these plant.^{6,7}

Aquatic plants (macrophytes) are important component of wetlands⁸, which have the special ability of adaption to the constant contact with surface water and groundwater⁹. Macrophytes have thin outer tissues and aerenchyma, through which air is distributed to the parts of plant below the surface of the water. Macrophytes vegetation has been used in wetlands to treat wastewater by uptake nutrients and heavy metals¹⁰. Macrophytes have a metabolic role in wastewater treatment due to their

potential to release oxygen into the rhizosphere which helps in nitrification and direct uptake of nutrients¹¹ as in *Typha* spp. Which has a considerably high nutrient uptake capacity¹².

Aquatic macrophytes are appropriate for metal removal as they can tolerate adverse conditions and are easier to harvest¹³. Wetland plants are divided into emergent, submerged and floating types. The emergent plants are rooted in the soil with basal portions, and leaves, stems and reproductive organs are aerial¹⁴. The examples of emergent plants are *Phragmites australis*, *Typha domingensis*, *Typha latifolia*, *Phragmites karka*, *Juncus pallidus*, *Empodisma minus*, *Phalaris arundinacea*, *Scirpus cyperinus*, *Aster novae-angliae*, *Limonium carolinianum*, *Cephalanthus occidentalis* and *Rhizophora mangle*. Submerged plants are below the surface of water for their entire life cycle. The examples of submerged plants are *Ceratophyllum demersum*, *Vallisneria americana*, *Myriophyllum spicatum*, *Hydrilla verticillata*, *Heteranthera dubia*, etc. Submerged species provide more biomass for the uptake and sorption of the contaminants through phytoextraction. Submerged plants have the ability to accumulate metals in their tissues in comparison to rooted emergent plants. In floating plants, the leaves and stems float on the surface of water. The examples of floating plants include *Eichhornia crassipes*, *Pistia stratiotes*, *Salvinia herzogii*, *Wolffia columbiana*, *Lemna valdiviana*, *Nymphaea* spp., *Nuphar advena*, *Juncus effusus*, *Phyllanthus fluitans*, etc.¹⁴.

The common aquatic plant species (*Typha latifolia*, *Myriophyllum exalbescens*, *Potamogeton epihydrus*, *Sparganium angustifolium*, *Myriophyllum spicatum* and *Sparganium multipedunculatum*) have been used for aluminium (Al) removal¹⁵. Macrophytes such as Parrot feather (*Myriophyllum aquaticum*), creeping primrose (*Ludwigia palustris*), and water mint (*Mentha* sp.) have been used for removal of iron (Fe), zinc (Zn), copper (Cu), and mercury (Hg) from water¹⁶. The *L. minor* has been used for removing copper (Cu) and cadmium (Cd) from contaminated soils¹⁷.

Aim of the study:

The main aim of this research work was to assess the remediation potential of select macrophytes. This involved:

- i) Weekly sampling of macrophytes
- ii) Identification of macrophytes samples
- iii) Assessment of remediation potential of selected wetland plants.

Materials and Methods

Study Area

Sampling of wetland plants

Wetland plants were sampled from inlets and outlets of Jakkur Lake, Bengaluru on a weekly basis (0.25 m² area). The plant samples were collected in triplicates. Collected macrophyte samples were washed, labelled and the plant species were identified using standard morphological keys based on taxonomic literature.¹⁸

Remediation potential of wetland plants

All the dried plant samples were powdered using mortar and pestle, sieved (1 mm) to get fine powders and labelled properly. The powdered plant sample (0.5 g) were acid digested¹⁹ and analysed for six heavy metals viz., cadmium (Cd), chromium (Cr), copper (Cu), nickel (Ni), lead (Pb) and zinc (Zn) using reagent blanks and suitable standards using Flame Atomic Absorption Spectrophotometry (GBC Avanta version 1.31).

Results and Discussion

Diversity of the wetland plants

Total seven wetland plant species were collected from Jakkur Lake, Bengaluru. The identified plant species were *Typha* sp. (Figure 1), *Cyperus* sp. (Figure 2), *Ludwigia* sp. (Figure 3), *Pistia stratiotes* (Figure 4), *Polygonum* sp. (Figure 5), *Alternanthera philoxeroides* (Figure 6) and *Spirodela* sp. (Figure 7). Among these five are emergent (*Typha* sp., *Cyperus* sp., *Ludwigia* sp., *Alternanthera philoxeroides* and *Polygonum* sp.) and two are floating wetland plant species (*Spirodela* sp. and *Pistia stratiotes*).



Fig. 1. *Typha* sp.



Fig. 2. *Cyperus* sp.



Fig. 3. *Ludwigia* sp.



Fig. 4. *Pistia stratiotes*



Fig. 5. *Polygonum* sp.



Fig. 6. *Alternanthera philoxeroides*

Synthesis, Characterization and Biological Activity of Zirconium Metal Complexes Derived from Novel Salens



Fig. 7. *Spirodela* sp.

Remediation potential of the wetland plants

Copper replaces other co-factors in key enzymes and disrupts photosynthetic activity and other cellular processes²⁰. Lead toxicity causes swollen, bent, short and stubby roots²¹. Excess Zn will alter the physiological, ultra structural and biochemical parameters of the plants. Excess of Ni causes various physiological alterations and diverse toxicity symptoms such as chlorosis and necrosis in the plant species²². Copper concentration exceeding 20 mg/kg in the shoot is injurious, though copper is an essential element for growth²³. The remediation potential of the wetland plants collected on weekly basis is represented in Tables 1, 2, 3 and 4 respectively.

Table 1. Uptake of heavy metals by the wetland plants (sampling done on 17/5/2017)

Plant samples	*Inlet						*Outlet					
	Heavy metals (mg/kg)						Heavy metals (mg/kg)					
	Cu	Pb	Zn	Ni	Cr	Cd	Cu	Pb	Zn	Ni	Cr	Cd
young <i>Typha</i> sp.	17.2	0.0	14.4	1.4	5.0	0.0	1.2	6.0	4.2	1.4	13.2	0.2
mature <i>Typha</i> sp.	-	-	-	-	-	-	0.0	3.2	3.2	0.6	10.0	0.0
young <i>Alternanthera philoxeroides</i>	0.0	0.0	2.8	2.0	0.6	0.0	0.0	7.6	3.0	5.8	10.8	0.0
medium <i>Polygonum</i> sp.	1.4	5.2	9.0	2.4	12.0	0.8	-	-	-	-	-	-
mature <i>Polygonum</i> sp.	0.8	0.0	3.2	0.2	8.8	0.2	-	-	-	-	-	-
medium <i>Ludwigia</i> sp.							0.6	8.4	6.4	6.2	10.6	0.6
mature <i>Pistia stratiotes</i>	-	-	-	-	-	-	2.6	8.2	7.0	6.2	9.6	0.2
<i>Spirodela</i> sp.	-	-	-	-	-	-	0.8	7.6	8.8	6.8	10.6	0.0
Normal range ²⁴	1-5	0.2-20	1-400	0.02-5	0.03-14	0.1-2.4						
Critical range ²⁴	5-30	30-300	100-400	10-100	5-30	5-30						
Threshold effect level ²⁵	35.7	18.0	123.0	35.0	37.3	0.596						



Table 2. Uptake of heavy metals by the wetland plants (sampling done on 30/5/2017)

Plant samples	*Inlet						*Outlet					
	Heavy metals (mg/kg)						Heavy metals (mg/kg)					
	Cu	Pb	Zn	Ni	Cr	Cd	Cu	Pb	Zn	Ni	Cr	Cd
young <i>Typha</i> sp. shoot	1.4	9.0	9.4	10.4	11.2	0.4	0.0	7.8	5.4	5.8	8.2	0.4
medium <i>Typha</i> sp. shoot	-	-	-	-	-	-	0.6	9.8	6.4	9.0	10.2	0.2
mature <i>Typha</i> sp. shoot	0.0	8.0	7.6	10.0	14.0	0.6	0.0	7.2	5.2	8.0	12.6	0.0
mature <i>Alternanthera philoxeroides</i>	-	-	-	-	-	-	0.6	8.8	10.2	5.6	10.0	0.4
mature <i>Pistia stratiotes</i>	-	-	-	-	-	-	1.2	5.0	8.8	6.0	7.0	1.6
<i>Spirodela</i> sp.	-	-	-	-	-	-	0.0	2.8	4.0	0.0	13.6	1.0
Normal range	1-5	0.2-20	1-400	0.02-5	0.03-14	0.1-2.4						
Critical range	5-30	30-300	100-400	10-100	5-30	5-30						
Threshold limit	35.7	18.0	123.0	35.0	37.3	0.596						

Table 3. Uptake of heavy metals by the wetland plants (sampling done on 6/6/2017)

Plant samples	*Inlet						*Outlet					
	Heavy metals (mg/kg)						Heavy metals (mg/kg)					
	Cu	Pb	Zn	Ni	Cr	Cd	Cu	Pb	Zn	Ni	Cr	Cd
medium <i>Typha</i> sp.	14.4	7.6	16.4	1.8	6.2	0.6	-	-	-	-	-	-
mature <i>Typha</i> sp.	2.0	4.6	5.2	4	6.0	0.8	1.0	13.6	10.4	0.0	2	0.2
mature <i>Alternanthera philoxeroides</i>	1.4	2.4	6.6	3.2	5.2	1.8	0.8	10.4	8.8	0.0	19.6	1.0
medium <i>Ludwigia</i> sp.	-	-	-	-	-	-	2.4	12.2	7.6	0.0	4.0	0.2
Normal range	1-5	0.2-20	1-400	0.02-5	0.03-14	0.1-2.4						
Critical range	5-30	30-300	100-400	10-100	5-30	5-30						
Threshold limit	35.7	18.0	123.0	35.0	37.3	0.596						

Metal uptake by wetland plants during the 1st week

The uptake of Cu by young *Typha* sp. from inlet exceeded the normal range and uptake of Ni by young *Alternanthera philoxeroides*, medium *Ludwigia* sp., mature *Pistia stratiotes* and *Spirodela* sp. from the outlet exceeded the normal range. (Table 1)

Metal uptake during the 2nd week

The uptake of Ni by young and mature *Typha* sp. shoot from inlet exceeded the normal range and uptake of Ni by young and mature *Typha* sp. shoot, mature *Alternanthera philoxeroides* and mature *Pistia stratiotes* from the outlet exceeded the normal range. (Table 2)

Synthesis, Characterization and Biological Activity of Zirconium Metal Complexes Derived from Novel Salens

Table 4. Uptake of heavy metals by the wetland plants (sampling done on 13/6/2017)

Plant samples	*Inlet						*Outlet					
	Heavy metals (mg/kg)						Heavy metals (mg/kg)					
	Cu	Pb	Zn	Ni	Cr	Cd	Cu	Pb	Zn	Ni	Cr	Cd
medium <i>Typha</i> sp.	2.0	5.8	7.6	1.8	8.8	0.8	-	-	-	-	-	-
mature <i>Typha</i> sp.	1.6	5.2	6.8	2.2	9.0	0.6	1.8	12.2	23	3.0	5.4	1.4
young <i>Alternanthera philoxeroides</i>	2.2	9.8	8.6	1.4	10.2	0.2	-	-	-	-	-	-
medium <i>Alternanthera philoxeroides</i>	-	-	-	-	-	-	0.4	0.0	3.6	2.4	4.6	0.8
medium <i>Ludwigia</i> sp.	-	-	-	-	-	-	3.2	3.6	7.8	3.4	9.6	0.6
Normal range	1-5	0.2-20	1-400	0.02-5	0.03-14	0.1-2.4						
Critical range	5-30	30-300	100-400	10-100	5-30	5-30						
Threshold limit	35.7	18.0	123.0	35.0	37.3	0.596						

*Inlet and outlet are the sampling points at Jakkur Lake, Bengaluru.

C - Carbon; N - Nitrogen; P - Phosphorus; Cd - Cadmium; Cr - Chromium; Ni - Nickel; Zn - Zinc; Cu - Copper; Pb - Lead

Metal uptake during the 3rd week

The uptake of Cu by medium *Typha* sp. from inlet exceeded the normal range and uptake of Cr by mature *Alternanthera philoxeroides* from the outlet exceeded the normal range. (Table 3)

Metal uptake by wetland plants during the 4th week

The uptake of heavy metals by the macrophytes during the 4th week was within the normal range. (Table 4)

Wetland plants at storm water detention ponds have showed the ability for the uptake of nutrients and heavy metals²⁶. *Eichhornia* sp. uptake heavy metals Pb, Cr, Zn, Mn, and Cu to a large extent from the wastewater²⁷. *Ludwigia natans* is useful for remediation of Cd from contaminated water²⁸. *Typha angustifolia* has been reported to assist remediation of various heavy metals (Cu, Pb, Ni, Fe, Mn, and Zn) and it has been found that the plant removes heavy metals from industrial wastewater containing metal, melanoidin, and phenol compounds²⁹. *Spirodela* sp. has high uptake capacity of Cr, Zn and Pb³⁰. The analyses highlights the use of plants for

remediation as an environmental friendly approach because of their high efficiency in metal removal³¹. Removal of toxic metals by wetland plants viz., *Phragmites australis*, *Juncus effusus* and *Iris pseudacorus*³². *Carex pseudocyperus* and *C. riparia* have been reported to be effective for the removal of metals from water³³. The heavy metal accumulation potential of wetland plants has been studied³⁴⁻³⁸. The spatial pattern of heavy metal accumulation in the sediments and macrophytes of Bellandur wetland, Bangalore, India show higher accumulation of all metals except for chromium in *Typha angustata*³⁵. The bioremediation potential of aquatic macrophytes in Jakkur wetland, Bangalore, India is evident with the higher concentration of lead, zinc, nickel and chromium in *Typha angustata* at inlet. Nickel above normal range was observed in *Typha angustata* at inlet and in all plant species at outlet³⁶⁻³⁸. Similarly, there is a report on heavy metals in biotic and abiotic components of Varthur wetlands, Bangalore, India^{37, 39}. The assessment of bioconcentration and translocation factors in macrophytes showed the select macrophytes are useful to remove heavy metals^{37, 38}.



Conclusions

The phytoremediation approach is an eco-friendly solution for the treatment of contaminated soils and water using plants. Common plants in wetlands such as *Typha* sp., *Cyperus* sp., *Ludwigia* sp., *Pistia stratiotes*, *Polygonum* sp., *Alternanthera philoxeroides* and *Spirodela* sp., have good remediation potential due to their metal uptake ability. Macrophytes have the ability for the uptake of heavy metals such as Cd, Pb, Zn, Ni, Cr and Cd, which will be the low cost option in the remediation of polluted soil and water. The uptake of heavy metals by the macrophytes during the fourth week was within the normal range suggesting that they can be used as fodder. Use of select local macrophyte species in the constructed wetland systems would aid in remediation through removal of contaminants (nutrients and heavy metals) from domestic and industrial wastewaters. Ensuring zero discharge from industries would be the best option toward ensuring heavy metals not getting into biotic food chain. Growing macrophytes in the flood plains of water bodies in rural area helps in treating agriculture run-off containing nutrients and heavy metals.

References

1. Vardhan K., Kumar P. and Panda R., 2019, *Journal of Molecular Liquids*, **290**.
2. Khulbe C. and Matsuura T., 2018, *Applied Water Science*, **8**, 19.
3. Azubuike C., Chikere B. and Okpokwasili C., 2016, *World Journal of Microbiology and Biotechnology*, **32**, 180.
4. Pande V., Pandey C., Sati D., Pande V. and Samant M., 2020, *Environmental Sustainability*, **3**, 91-103.
5. Etim E., 2012, *International Journal of Environment and Bioenergy*, **2**, 120-136.
6. Lee H., 2013, *Biotechnology and Bioprocess Engineering*, **18**, 431-439.
7. Wei Z., Van Le Q., Peng W., Yang Y., Yang H., Gu H., Lam S. and Sonne C., 2021, *Journal of Hazardous Materials*, **403**.
8. Gupta P., Roy S. and Mahindrakar A., 2012, *Resources and Environment*, **2**, 202-215.
9. Kochi L., Freitas P., Maranhão L., Juneau P. and Gomes M., 2020, *Sustainability*, **12**, 9202.
10. Galal M. and Farahat A., 2015, *Environmental Monitoring and Assessment*, **187**, 701.
11. Guo-feng L., Zhang lu., Cheng-xin F., Shi-qun H., Jun H. and Paerl H., 2014, *Journal of Pollution Effects and Control*, **2**, 113.
12. Maddison M., Muring T., Remm K., Lesta M. and Mander U., 2009, *Ecological Engineering*, **35**, 258-264.
13. Jha P., Samal A., Santra S. and Dewanji A., 2016, *American Journal of Plant Sciences*, **7**, 2112-2137.
14. Herath I. and Vithanage M., 2015, Phytoremediation in constructed wetlands. In: Phytoremediation: Management of environmental contaminants, Ansari A., Gill S., Gill R., Lanza R. and Newman L (Eds.), Vol 2, Springer International Publishing, Switzerland.
15. Gallon C., Munger C., Premont S. and Campbell P., 2004, *Water, Air, & Soil Pollution*, **153**, 135-155.
16. Kamal M., Ghaly A., Mahmoud N. and Cote R., 2004, *Environment International*, **29**, 1029-1039.
17. Hou W., Chen X., Song G., Wang Q. and Chang C., 2007, *Plant Physiology and Biochemistry*, **45**, 62-69.
18. Cook C., 1996, Aquatic and wetland plants of India, Oxford New York, Oxford University Press.

Synthesis, Characterization and Biological Activity of Zirconium Metal Complexes Derived from Novel Salens

19. APHA, 1995, Standard Methods for the examination of water and wastewater, 19th ed., American Public Health Association, Washington, DC.
20. Arguello J., Raimunda D. and Padilla-Benavides T., 2013, *Frontiers in Cellular and Infection Microbiology*, **3**, 1-14.
21. Nas F. and Ali M., 2018, *MOJ Ecology and Environmental Sciences*, **3**, 265-268.
22. Gill M., 2014, *International Journal of Advanced Research*, **2**, 1043-1055.
23. Singh S., Parihar P., Singh R., Singh V. and Prasad S., 2016, *Frontiers in Plant Science*, **6**, 1-36.
24. Maiti S.K., 2003, Handbook of Methods in Environmental Studies: Air, noise, soil and overburden analysis, Vol 2, ABD Pub, Oxford Book Co, India.
25. MacDonald D., Ingersoll C. and Berger A., 2000, *Archives of Environmental Contamination Toxicology*, **39**, 20-31.
26. Istenic D., Arias C. and Brix H., 2011, The Proceedings Strepow Workshop, Andrevlje-Novi Sad, Serbia, p.111-117.
27. Tiwari S., Dixit S. and Verma N., 2007, *Environmental Monitoring Assessment*, **129**, 253-256.
28. Marbaniang D. and Chaturvedi S., 2014, *Keanean Journal of Science*, **2**, 29-34.
29. Chandra R. and Yadav S., 2010, *Ecological Engineering*, **36**, 1277-1284.
30. Ensley D., 2000, Rational for use of phytore-mediation. In: Phytoremediation of toxic metals: Using plants to clean- up the environment, Raskin I. and Ensley D (eds), New York, John Wiley & Sons, Inc, p.3-12.
31. Akbarzadeh A., Jamshidi S. and Vakhshouri M., 2015, *Pollution*, **1**, 1-8.
32. Perez-Sirvent C., Hernandez-Perez C., Martinez-Sánchez M., Lorenzo M. and Bech J., 2017, *Journal of Soils and Sediments*, **17**, 1384-1393.
33. Schuck M. and Greger M., 2020, *International Journal of Environment Research and Public Health*, **17**, 4623.
34. Wu H., Zhang J., Ngo H., Guo W., Hu Z., Liang S., Fan J. and Liu H., 2015, *Bioresource Technology*, **175**, 594-601.
35. Ramachandra T., Sudarshan B., Mahesh K. and Vinay S., 2018, *Journal of Environmental Management*, **206**, 1204-1210.
36. Sudarshan B., Mahesh M. and Ramachandra T., 2019, *Indian Journal of Environmental Protection*, **39**, 594-601.
37. Ramachandra T.V., Sudarshan B., Shivamurthy V., Asulabha S. and Varghese S., 2020, *SN Applied Sciences*, **2**, 3228-3238.
38. Bharath H. Aithal and Ramachandra T. V., 2016, *J. Indian Soc. Remote Sens.*, **44(4)**, 617-633.
39. Ahalya, N., Kanamadi R.D. and Ramachandra T.V., 2007, *Journal of Enviro. Bio.*, **28(4)**, 765-769.